# **Development of Peptide-Based Quorum Sensing Modulators that Attenuate Streptococcus** pneumoniae Pathogenicity

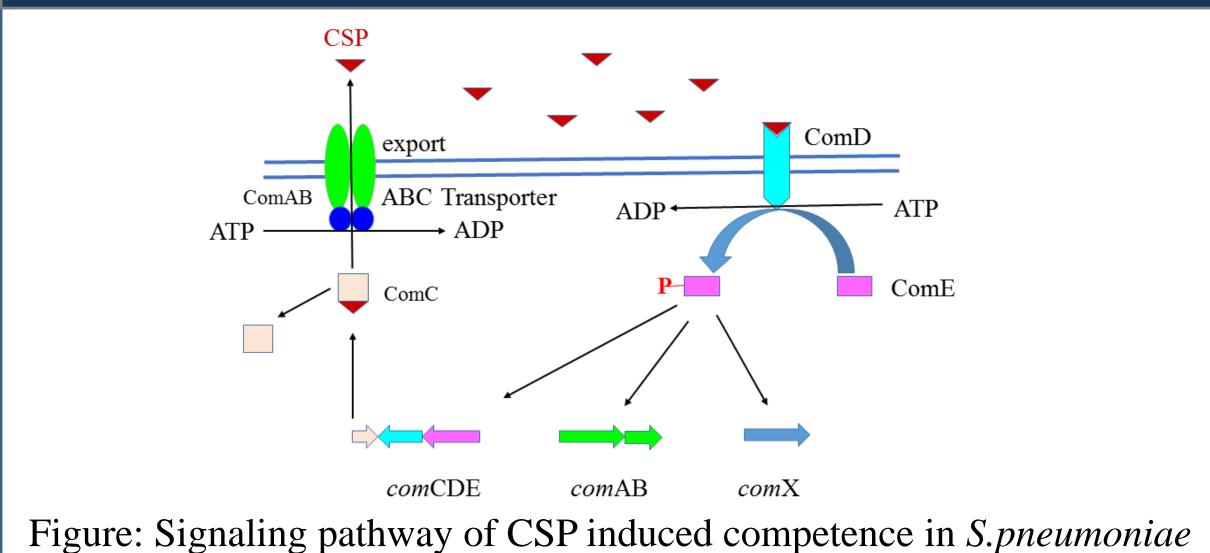
### Introduction

The discovery of bacterial communication changed our general perception of microscopic organisms. Bacteria use signaling molecules (sometimes referred to as pheromones) released into the environment as a chemically-based language which allows for both intraspecies and interspecies communication.<sup>1</sup> The release and detection of the signaling molecule(s) allows bacteria to measure the number of bacteria within a population. This phenomenon is known as quorum sensing (QS).<sup>2</sup> QS is a process in which the concentration of signaling molecules enables a single cell to sense the overall bacterial population and allows bacteria to change their behavior once a threshold concentration is reached. When the threshold signaling molecule concentration is reached, bacteria coordinate their behavior through synchronization and the establishment of group behaviors such as the formation of biofilms, induction of competence, and release of virulence factors (REF).<sup>2,3</sup> This cooperation is important for pathogenic bacteria to communicate together in order to establish infection and to escape the host's immune response.<sup>3</sup> Bacteria are omnipresent in nature, different bacterial species use unique molecules to and communicate with each other

## Streptococcus pneumoniae

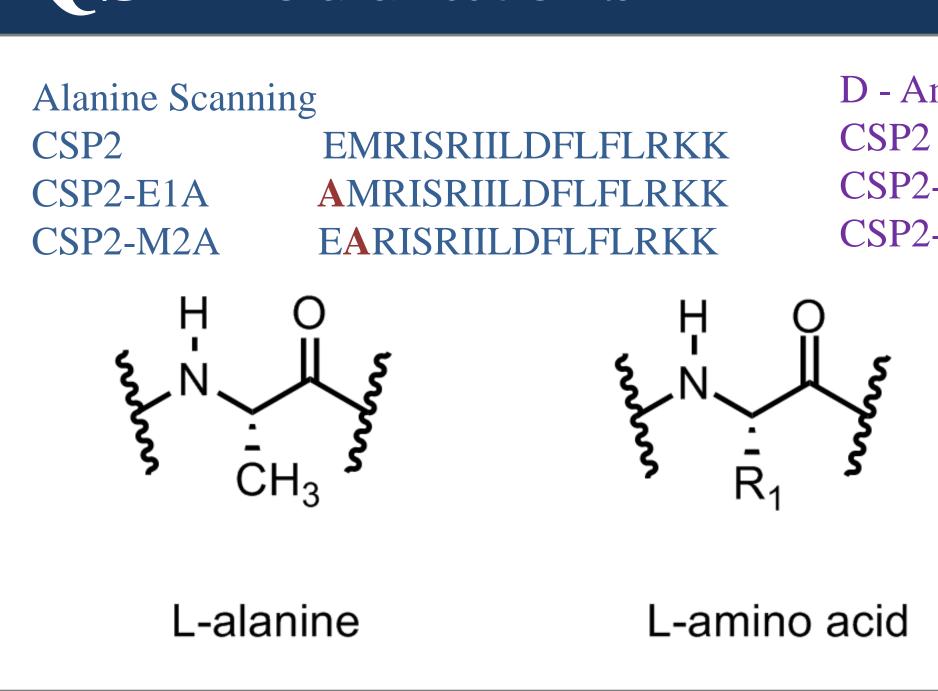
- Streptococcus pneumoniae is a gram-positive bacteria that inhabits the nasopharynx of human and cause numerous disease.
- $\succ$  S. pneumoniae acquires antibiotic resistant genes through genetic transformation
- $\succ$  Competence in S. pneumoniae is initiated by the competence stimulating peptide (CSP)-mediated quorum sensing (QS) circuit
- $\succ$  S. pneumoniae strains can be divided into two main specificity groups, Group 1 and Group 2, based on the CSP signal (CSP1 or CSP2) and associated receptor (ComD1 or ComD2) they produce.<sup>4</sup>
- ▶ In this study we systematically altered the sequence of CSP2 to find analogs that can modulate the QS response and attenuate the pathogenicity of S. pneumoniae.

## **CSP** Pathway



Bimal Koirala, Naiya R. Phillips, Sally R. Hamry, and Yftah Tal-Gan Department of Chemistry, University of Nevada, Reno

## **QS** Modulators



## **Structure Activity Relationship of CSP2**

| Name      | EC <sub>50</sub> /IC <sub>50</sub> (nM)<br>ComD-1 | ComD-2 | Name            |
|-----------|---|--------|-----------------|
|           |   |        | CSP2            |
| CSP2-E1A  | >1000   | >1000  | CSP2-e1         |
| CSP2-M2A  | <sup>a</sup>                                      | 897    | CSP2-m2         |
| CSP2-R3A  |   |        | CSP2-r3         |
| CSP2-I4A  |   | 170    | CSP2-i4         |
| CSP2-S5A  |   | 148    | CSP2-s5         |
| CSP2-R6A  | >1000   | 116/   | CSP2-r6         |
| CSP2-I7A  | >1000   | 250/   | CSP2-i7         |
| CSP2-I8A  |   |        | CSP2-i8         |
| CSP2-L9A  |   | 30.1   | CSP2-19         |
| CSP2-D10A | >1000   | 319    | <b>CSP2-d10</b> |
| CSP2-F11A |   | 164    | CSP2-f11        |
| CSP2-L12A |   | >1,000 | CSP2-112        |
| CSP2-F13A |   | 505    | CSP2-f13        |
| CSP2-L14A |   | 175    | CSP2-114        |
| CSP2-R15A |   | 24.3   | CSP2-r15        |
| CSP2-K16A |   | 80.5   | CSP2-k16        |
| CSP2-K17A |   |        | CSP2-k17        |
|           |   |        |                 |

Table 1:  $EC_{50}/IC_{50}$  values of the alanine and D-amino acid scan analogs of CSP2 against ComD2 receptor.

<sup>a</sup> EC50/IC50 not determined due to the low induction, the analog shows in initial screening.

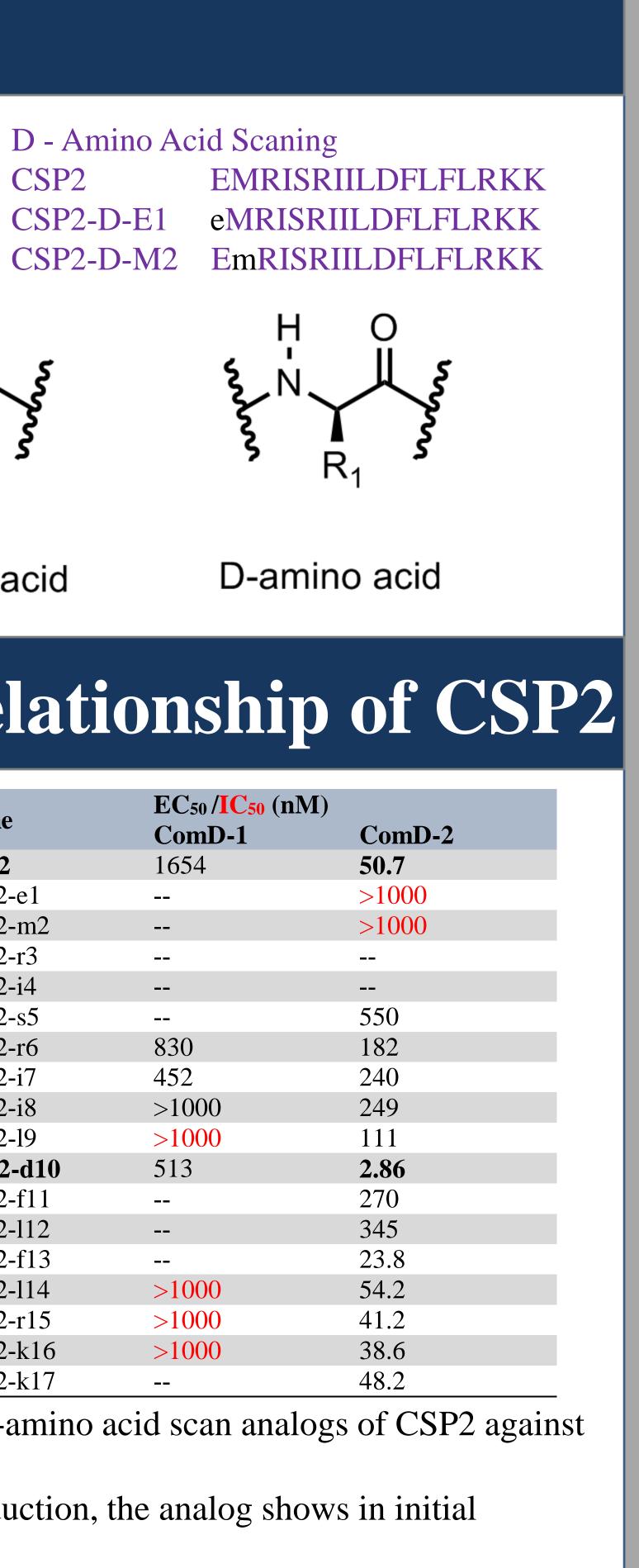
| $\begin{array}{c ccccccccccccccccccccccccccccccccccc$ |  |
|---|--|
|---|--|

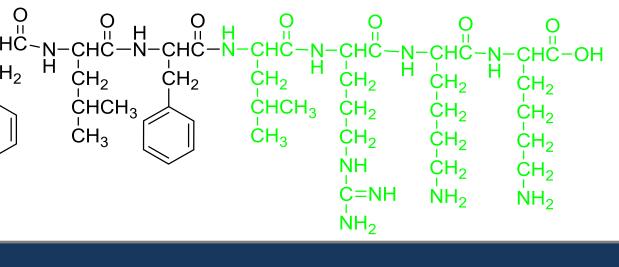
### Second Generation CSP2

| Name                  | EC <sub>50</sub> /IC <sub>50</sub> (nM)<br>ComD-2 |
|-----------------------|---|
| CSP2                  | 50.7  |
| CSP2-des-K17          | 44.4  |
| CSP2-desK17K16        | 21.8  |
| CSP2- desK17K16R15    | 42.3  |
| CSP2- desK17K16R15L14 | 77.8  |
| CSP2-E1Ad10           | 56.5  |

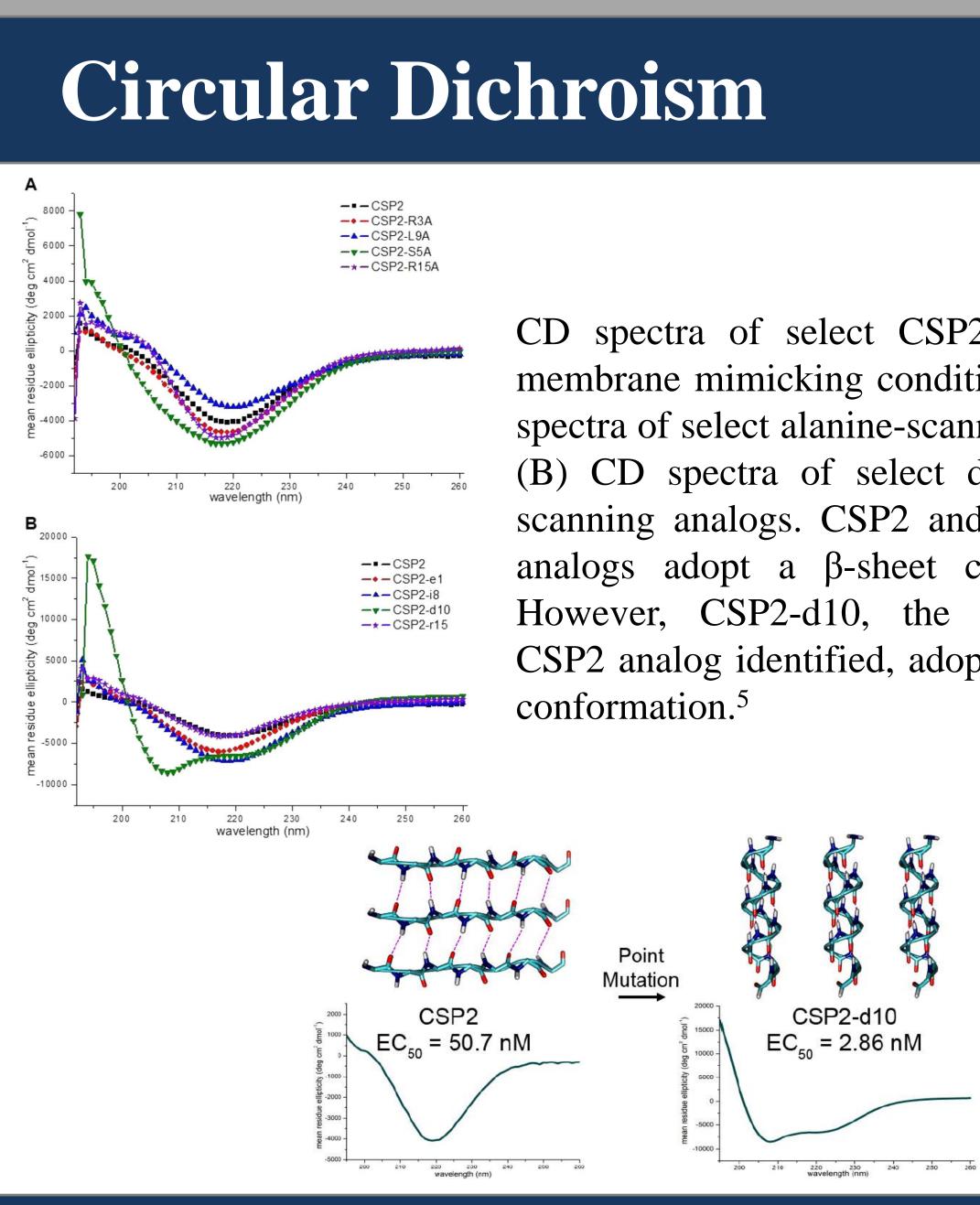
 $\blacktriangleright$  We made more than 15 second generation analogs  $\succ$  We were able to rational design to make 1<sup>st</sup> good inhibitor(CSP2-E1Ad10).<sup>5</sup>

- $\succ$  C- terminus is dispensable
- rational analogs.





 $\succ$  This result gives us a new window to make more



### Summary

- amino acid with receptor.
- respectively.
- binding to the ComD2 receptor
- $\succ$  Suggests design of third generation peptides.

## References

## Acknowledgement

"The project described was supported by the Nevada INBRE through a grant from the National Institute of General Medical Science (P20GM103440) from the National Institutes of Health."

CD spectra of select CSP2 analogs in membrane mimicking conditions. (A) CD spectra of select alanine-scanning analogs. (B) CD spectra of select d-amino acid scanning analogs. CSP2 and most of its analogs adopt a  $\beta$ -sheet conformation. However, CSP2-d10, the most active CSP2 analog identified, adopts an  $\alpha$ -helix

> Alanine and D-amino acid scanning shows correlation of specific

 $\succ$  N-terminus is very critical for activating the receptor.  $\succ$  CSP2-d10 and CSP2-E1Ad10 are able to turn on and off QS

 $\succ$  The CD analysis results suggest that  $\alpha$ -helix is required for effective

1) Miller, B. M.; Bassler, L.B. Annu. Rev. Microbiol. 2001, 55, 165. 2) Dunny, M. G. Annu. Rev. Microbiol. 1997, 51, 527. 3) Waters, C. M.; Bassler, L. B. Annu. Rev. Cell Dev. Biol. 2005, 21, 319 4) Zhu, L.; Lau, W. G. *PLoS Pathog.* **2012,** *7*, 1. 5) Yang, Y.; Koirala, B.; Sanchez, L.; Phillips, R. N.; Hamry, R. S.; Tal-Gan, Y. ACS Chem. Biol. 2017, 12, 1141-1151